SECONDARY METABOLITES AS POTENTIAL THERAPEUTIC AGENTS AGAINST PARASITIC INFECTIONS

Abstract

In the context of global health, parasitic diseases continue to be a significant challenge, particularly developing countries with limited resources. The emergence of drug-resistant parasites and the undesirable side effects associated with conventional treatments have triggered the search for alternative therapeutic strategies. Secondary metabolites, which are compounds synthesized natural by organisms for ecological reasons, offer great promise as potential effective agents against parasitic infections. This book chapter aims to present a comprehensive summary of recent research on secondary metabolites with anti- parasitic properties, emphasizing their mechanisms of action, efficacy, and potential applications in the treatment of parasitic diseases.

Keywords: parasitic infections, metabolites, phytocompounds, therapeutic agents.

Authors

Shabir Ahmad Rather

Department of Zoology School of Biosciences and Biotechnology Baba Ghulam Shah Badshah University Rajouri, Jammu & Kashmir,India.

Pooja Bharti

Department of Zoology University of Kashmir Srinagar, Jammu & Kashmir, India.

M. A. Hannan Khan

Department of Zoology School of Biosciences and Biotechnology Baba Ghulam Shah Badshah University Rajouri, Jammu & Kashmir, India.

Zahoor Ahmad Wani

Division of Veterinary Parasitology SKUAST-K Shuhama, Jammu & Kashmir, India. ahmadzahoor11@gmail.com

INTRODUCTION

I. SECONDARY METABOLITES AS THERAPEUTIC AGENTS

Throughout their life cycle, plants create secondary metabolites (SMBs). These SMBs are produced from primary substances that act as their precursors, including lipids, proteins, carbohydrates, and nucleic acids (Saltveit 2017). The range of organic compounds that fall under SMBs include terpenes, resins, alkaloids, flavonoids, and phenolic compounds, as depicted in **Figure 1**. These secondary metabolites contribute to the color, flavor, and fragrance of plants (Shitan 2016; Bansal et al., 2017; Symeonidou et al., 2018; Chanda and Ramchandra 2019). Notably, essential oils, polyphenols, and glycosidic glucosinolates extracted from various plant species, especially medicinal and aromatic plants, have demonstrated significant potential in combating diseases caused by various pathogens, owing to their remarkable antimicrobial activity.

II. SOURCES OF SECONDARY METABOLITES WITH ANTI-PARASITIC ACTIVITY

Numerous secondary metabolites have been sourced from plants, microorganisms, and fungi. For instance, alkaloids like morphine and quinine have been extracted from plants and employed as potent pain relievers and antimalarial medications, respectively. Additional examples encompass taxol, derived from the Pacific yew tree, utilized in treating different cancers, and artemisinin, obtained from *Artemisia annua*, known for its effectiveness against malaria. Secondary metabolites remain a valuable reservoir of new drug candidates, and their investigation offers great potential for the advancement of novel therapeutics to enhance human and animal well- being.

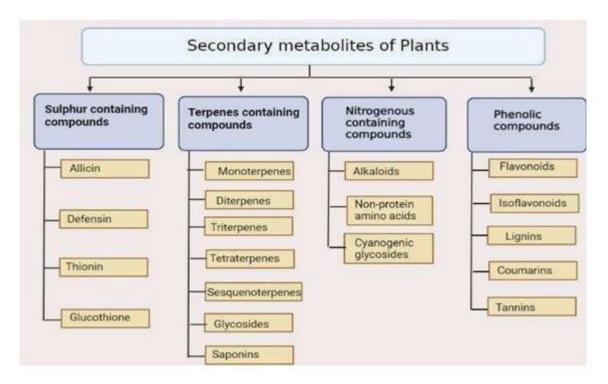


Figure 1: Classification of Secondary Metabolites of Plants.

Several classes of secondary metabolites have demonstrated anti-parasitic potential. Alkaloids, terpenoids, flavonoids, quinones, and polyphenols are discussed in the following table (**Table 1**).

Table 1: Structural and Functional Properties of Secondary Metabolites

1. Terpenes	Terpenes consist of distinct hydrocarbons formed by arranging various 5-carbon isoprene units. Within this category, glycosides and saponins are encompassed, their classification being determined by their respective structural characteristics.
2. Glycosides	Glycosides arise as substitutes for acetals, resulting from the interaction between monosaccharides and alcohol with the aid of an acidic catalyst.
3. Saponins	Saponins, often represented as SN, consist of either a triterpene or steroidal aglycone coupled with one or several chains of sugar. They are found in more than 100 plant families and act as active ingredients. Saponins are predominantly present in monocotyledons and less frequently in dicotyledons.
4. Phenolic compounds	Phenolic compounds: Phenolic compounds (PC) are characterized by a functional heterogeneous group with an aromatic ring. They are metabolites formed by the condensation of acetate units and include flavonoids, isoflavonoids, and tannins.
5. Flavonoids	Flavonoids (FN) are low molecular weight secondary metabolites that serve various functions such as defense mechanisms, safeguarding against UV radiation, inhibiting auxin transport, displaying allelopathic effects, and contributing to the pigmentation of flowers.
6. Tannins	Tannins (TN) are polyphenolic compounds soluble in water, with the ability to cause proteins like gelatin to precipitate. These compounds are present in a wide range of plants, encompassing both trees and herbs.

Table 2: Mechanism of Action of Secondary Metabolites to Control Parasitic Infection

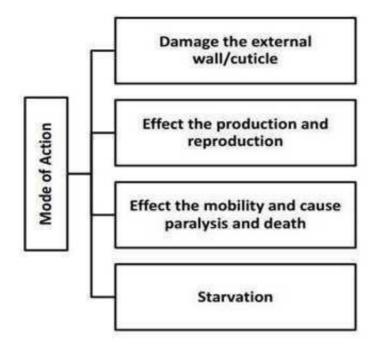


Table 3: Impact of Parasitism on Different Hosts.

Class	Site of infection	Host	Parasites	Consequence of parasitism
	Blood	Human	Plasmodium	Anemia
Protozoa		Human	Babesia	
		Bovine	Babesia	
			divergens	Fever, chills,
	Blood	Ticks		haemolytic
		(Intermediate	Babesia mircoti	anemia
		host)		
		White- footed		
		mouse;		
		Peromyscus		
		leucopus		
		(Primary host) and <i>Ixodes</i>		
		(Secondary host)		
		11081)	Haemonchus	Anemia
Nematode	Abomasum	Sheep, goat	contortus	Reduced feed intake
			Teladorsagia	
			circumcincta	

	Small intestine	Sheep, goat	Nematodirus battus Trichostrongylus colubriformis	Reduced feed efficiency
	Abomasum	Cattle	Haemonchus placei Ostertagia ostertagi	Reduced feed intake
	Small intestine	Cattle	Cooperia oncophora Cooperia punctata	Reduced feed efficiency
Cestode	Small intestine	Human	Taenia solium Taenia saginata	Abdominal distress, anorexia, diarrhea, dyspepsia Nausea, appetite loss and weight loss
	Small intestine Small	Canids Rodents and	Echinococcus granulosus	Pulmonary and liver cyst formation Headache,
	intestine	human (intermediate host)	Hymenolepis nana	abdominal pain, diarrhoea

III. SECONDARY METABOLITES AGAINST PROTOZOA

1. *Protozoan diseases:* Protozoan diseases pose significant economic and health challenges worldwide. These diseases, which affect both humans and livestock, include malaria, trypanosomiasis, chagas disease, leishmaniasis, and toxoplasmosis (Capela et al., 2019). The increasing prevalence of drug- resistant strains and the limited development of new drugs with innovative approaches have reduced the efficacy of current treatments for these diseases. Protozoa are microscopic, single-celled eukaryotic organisms found worldwide, with over 65,000 described species, many of which are free-living parasites. These organisms exhibit complex internal structures and intricate metabolic activities (Shanan et al., 2015). The life cycle of protozoan parasites involves feeding trophozoites, which can be either intracellular or extracellular. Parasite transmission occurs through various modes, including the fecal-oral route, vector-borne transmission, and predatory-prey interactions (Imam et al., 2009; Antonovics et al., 2017). Additionally, protozoan parasites undergo a dormant stage, allowing them to survive in extreme conditions without oxygen and nutrients for extended periods.

- **2.** Giardiasis: Giardia is a common flagellate of the intestine of mammals (Gardner and Hill, 2001). Giardia is a newly prominent parasite affecting livestock and diverse wildlife categories, spanning from birds to amphibians worldwide. Within the genus Giardia, numerous species share resemblances in terms of structure and possess multiple affiliations with different hosts. The infected animal may act as zoonotic potential of infection of Giardia. The transmission may occur from an infected animal to a healthy animal or human. However, the direct transmission of Giardia from infected animal to healthy animal or human still needs to be explored. The escalation of this ailment to a global health concern is attributed to the development of resistance to commonly employed drugs, compelling the exploration of plant-derived remedies for various parasitic infections. Pippali Rasayana, an Ayurvedic herbal remedy, is concocted from Piper longum (Pippali) and B. monosperma (Palash), utilizing the ash of the stem, root, flower, and leaves of B. monosperma. This preparation has displayed substantial efficacy against giardiosis, exhibiting a recovery rate of up to 98% from infections (Agrawal et al., 1997). In- vitro assessments have identified notable herbal contenders against G. lamblia, including Allium sativum, Artemisia sieberi, and Chenopodium botrys, demonstrating maximum impact at a concentration of 0.1% µg/ml. Helianthemum glomeratum methanolic extract exhibited exceptional effectiveness with an ED50 value of 0.125 mg/kg against giardiasis in animal models. Lavender's hydroalcoholic extract at a dose of 400 mg/ml exhibited potency against Giardia lamblia. Ferula assa-foetida's alcoholic extract displayed efficacy of 37% at 20 mg/ml concentration, reaching 100% by the 4th hour against Giardia lamblia. In Balb/C mice, Tanacetum parthenium's chloroformextract at 100 mg/ml concentration proved effective against Giardia lamblia. Another notable contender, the crude extract from Agereatum conivzoides leaves, exhibited strong effectiveness against Giardia duodenalis trophozoites (Pintong et al., 2020). Furthermore, the potential of ethanolic extract from pomegranate peel was assessed against G. lamblia, revealing significant reduction in cysts and trophozoites within faecal matter and the intestines, respectively.
- **3.** *Toxoplasmosis: Toxoplasmosis*, a zoonotic disease caused by the obligate parasite *Toxoplasma*, is widely distributed worldwide. *T. gondii*, a single species in the genus Toxoplasma belonging to the phylum Apicomplexa, can be life-threatening in individuals with weakened immune systems. In recent times, there has been increasing focus on exploring alternative treatments for parasitic diseases, leading to numerous studies in this area. Fungi have the capacity to produce a diverse array of secondary metabolites (Costaet al., 2016), including peptides, alkaloids, terpenes, polyketides, quinones, sterols, and coumarins. These secondary metabolites from fungi have shown effectiveness against protozoan infections.
- **4.** *Malaria: Plasmodium*, a unicellular protozoan parasite transmitted through the bite of infected *Anopheles* mosquitoes, causes malaria, which is more prevalent in tropical and sub-tropical regions worldwide. The four common species responsible for malaria are *P. falciparum*, *P. vivax*, *P. malariae*, and *P. ovale*, with *P. falciparum* and *P. vivax* being the most common. In India, *Plasmodium vivax* is the primary cause of malarial infection (Anvikar et al., 2016). The World Health Organization (WHO) reports that the global malaria burden mainly originates from 15 countries in sub-Saharan Africa and the Indian subcontinent, with five of those countries, including India, accounting for half of all malaria cases (WHO, 2018).

Malaria treatment involves a range of available drugs like chloroquine, mefloquine, primaquine, and artemisinin. Yet, the rise of drug-resistant *Plasmodium* strains hampers their effectiveness significantly (Bahekar et al., 2013). Particularly, *Plasmodium*, notably *P. falciparum* causing tropical malaria, has demonstrated resistance to numerous synthetic medications. A remarkable stride in antimalarial drug development was the unveiling of artemisinin, a sesquiterpene lactone derived from *Artemisia annua* (Asteraceae). This compound has proven efficacy even against *P. falciparum* strains resistant to multiple drugs (Willcox, 2011; Efferth et al., 2011). Currently, derivatives of artemisinin created through partial synthesis, such as the water-soluble artesunate, are being used in clinical applications (Kuhn and Wang, 2008). Medicinal plants often harbor intricate combinations of secondary metabolites, and there's a belief that these compounds work together synergistically when present in extracts (Wink, 2008). For instance, combining epigallocatechin 3-gallate (EGCG), a common polyphenol found in green tea, with the saponin digitonin has demonstrated synergistic effects in diminishing the mobility and viability of *Plasmodium berghei* (Hellmann et al., 2010).

Numerous plants have been identified as sources of anti-malarial medicinal properties, with the genus Artemisia being particularly notable. Essential oils and biomolecules derived from these plants, such as artemisinin from *Artemisia annua*, have shown antimalarial activity. Additionally, *Cinchona officinalis* and related species from the Rubiaceae family were among the first used for developing anti-malarial drugs, with quinoline alkaloids like quinine being effective against different developmental stages of the malaria parasite. Combinations of *Allium sativum* with other plants have also been suggested for the effective treatment of *Plasmodium* (Alebie et al., 2017).

Furthermore, various compounds derived from fungi, such as sterosterin A from *Stereum ostera*, Aurisin A, G, and K from *Neonothopanus nimbi* and *Anthracophyllum* species, Eurochevalierine from *Eurotium chevalieri*, and Hirsutellone F from *Trichoderma* spp. have exhibited anti-plasmodial effects, particularly against *P. falciparum*.

5. Coccidiosis: Coccidiosis, a major parasitic disease in poultry caused by protozoan parasites of the genus Eimeria, has a significant impact on the growth and feed utilization of infected birds, leading to productivity loss. This poultry disease affects the epithelial lining of the intestine and is prevalent worldwide. Infected birds may display symptoms such as droopiness, pale comb, diarrhoea, and occasional blood in droppings, with high mortality rates observed in both chicks and adults. Current strategies for preventing and treating coccidiosis include the use of anticoccidial chemicals, vaccines, and natural products. Plant-based phytochemicals have emerged as a promising and safe alternative for managing coccidiosis. Garlic and its sulfur compounds, such as allicin, alliin, ajoene, diallyl sulfide, dithiin, and allylcysteine, have been reported to exhibit broad antimicrobial activities, effectively inhibiting the sporulation of Eimeria tenella in vitro (Alnassan et al., 2015; El-khatam et al., 2014; Pourali et al., 2014). Green tea extracts have also shown significant inhibition of coccidian oocyst sporulation, with selenium and polyphenolic compounds thought to be the active agents responsible for inactivating enzymes involved in sporulation. Studies on the extract of Garcinia kola demonstrated its anticoccidial activity against experimental Eimeria tenella infection in broiler chickens (Shetshak et al., 2020). Azadirachta indica, commonly known as neem,

contains various compounds, including limonoids and protolimonoids, which are believed to influence Eimeria's life cycle (Biswas et al., 2002; Koul et al., 2006). In comparison to salinomycin sodium, neem fruit added to broiler diets showed efficient repression of coccidiosis (Tipu et al., 2002).

Furthermore, essential oils (EO) derived from aromatic plants and their constituents have displayed antimicrobial properties against bacteria and fungi in both laboratory experiments and living organisms (Rhayour et al., 2003, Chami et al., 2004, 2005, and Bennis et al., 2004). Recent research conducted by Remmal et al. (2011) demonstrated the efficacy of essential oils in destroying *Eimeria o*ocysts in laboratory settings. The major components of EO, including carvacrol, carvone, isopulegol, thymol, and eugenol, were tested for their ability to eliminate *Eimeria* oocysts in a dose and time-dependent manner, as indicated by the release of substances absorbing at 273 nm.

6. *Trichomoniasis:* Trichomoniasis, caused by *Trichomonas vaginalis*, is a prevalent sexually-transmitted protozoan infection that primarily affects women but also affects men to a lesser extent. Globally, approximately 170 million people are infected, and this infection is often linked to HIV and cervical cancer (Gehrig et al., 2009). Due to the emergence of drug-resistant *Trichomonas* strains, there is a demand for new chemical treatments. Natural products, such as alkaloids like berberine, dibenzofurans, anthraquinones, polyacetylenes, saponins, and diterpenes, have been investigated as potential alternative therapies (Gehrig et al., 2009).

IV. SECONDARY METABOLITES AGAINST NEMATODES (LYMPHATIC FILARIASIS)

Lymphatic filariasis (LF) poses a significant global health challenge, impacting approximately 120 million individuals in 72 countries. The disease is caused by nematode parasites Wuchereria bancrofti, Brugia malayi, and Brugia timori, which are transmitted through mosquito bites from species like Culex, Anopheles, Aedes, and Mansonia (Lourens et al., 2019). LF is closely associated with lymphedema, commonly known as elephantiasis, resulting in swelling of limbs, breasts, and genitals. In response, the World Health Organization (WHO) has launched the Global Program to Eliminate Lymphatic Filariasis (GPELF) with the goal of curbing the spread of infection, eradicating LF, and alleviating the suffering of affected individuals. Current treatment strategies involve mass drug administration of drugs like albendazole, diethylcarbamazine (DEC), and ivermectin, which target the microfilariae stages but are inadequate in eliminating the adult worms. Consequently, research has explored the potential of herbal plants for their anti-filarial properties. Notably, Oldenlandia herbacea, Buteamon sperma, Streblus asper, and Adansonia digitata have exhibited significant anti-filarial effects both in laboratory studies and living organisms (Singh and Singh, 1987; Chatterjee et al., 1992; Ibrahim et al., 2014), and their use may aid in regulating inflammation associated with LF pathologies.

1. Ascaridiasis in Poultry: *Ascaridia galli*, a parasitic nematode affecting poultry, presents a widespread and economically significant challenge on a global scale. Various anthelmintic agents, including piperazine, albendazole, levamisole, ivermectin, benzimidazole, and fenbendazole, have proven effective in managing *A. galli*

infections. Fenbendazole stands out due to its distinctive mode of action (Bazh and El-Bahy, 2013; Yazwinski et al., 2013; Umar et al., 2018). Certain plants contain secondary metabolites (SMBs) like terpenes (glycosides and saponins), phenolic compounds (flavonoids and tannins), and nitrogen-containing compounds (alkaloids, cyanogenic glycosides, and non-protein amino acids). These compounds exhibit anthelmintic properties by influencing nematodes through various mechanisms (Zaman et al., 2020). Additionally, citrus peels housing Limonene, β -Pinene, α -Pinene, and Sabinene have displayed potential anthelmintic attributes against *A. galli* (Abdelqader et al., 2012).

2. Cysticercosis: Cysticercosis, a result of the pork tapeworm *Taenia solium*, has become a notable issue for public health and agriculture in underdeveloped areas across Latin America, Africa, and Asia. This concern arises particularly in regions where pigs are reared for consumption through traditional methods. This zoonotic ailment gives rise to cysts within both humans and pigs, ultimately causing epilepsy and fatalities in humans. Concurrently, it diminishes the market value of pigs and renders pork unfit for consumption. The transmission of *T. solium* occurs between humans and between humans and pigs, occasionally even exclusively among humans, resulting in the development of cysticercosis.

V. SECONDARY METABOLITES AGAINST TREMATODIASIS (SCHISTOSOMIASIS)

Schistosomiasis: Schistosomiasis is commonly managed through the administration of praziquantel, either as a standalone treatment or in conjunction with albendazole. Other therapeutic options encompass oxamniquine and antimalarial medications such as quinoline alkaloids and artemisinin, along with its derivatives. These treatments have been utilized for addressing the ailment (Mullner et al., 2011). Efforts against the vector snails, responsible for schistosomiasis transmission, namely *Oncomelania*, *Biomphalaria*, and *Bulinus*, have identified molluscicidal properties in various compounds. Anthraquinones present in *Rheum palmatum* and *Rumex dentatus* (Polygonaceae), phorbol esters derived from *Jatropha curcas* (Euphorbiaceae), and the broader category of saponins have displayed activity in this regard (Liu et al., 1997). Furthermore, Curcumin and its derivatives, sourced from Curcuma plants (Zingiberaceae), have demonstrated parasiticidal effects against *Schistosoma* parasites (Haddad et al., 2011).

Table 4: Active Phytocompounds for Treatment of Parasitic Diseases.

Family	Plant Name	Active Compounds	Parasite disease	Reference
Malvaceae	Adansonia digitata	Polysaccharide	Lymphatic filariasis	Ouedrago et al., 2020
Fabaceae	O	Flavonoids, saponins, tannins and Triterpenes	Lymphatic filariasis	Saleh <i>et al.</i> , 2021
Meliaceae	Azadirachta indica	Tetranotriterpenoids	Lymphatic filariasis	Mukherjee et al. 2019
Lamiaceae	Ocimum tenuiflorum	Essential oil	Lymphatic filariasis	Malebo et al.,2012

Lamiaceae	Ocimum	Essential oil	Lymphatic	Malebo et al., 2012
	gratissimum		filariasis	
Lamiaceae	Hyptis suaveolens	Essential oil	Lymphatic	Malebo et al.,2012
			filariasis	

Table 5: Action of Phytocompounds against Parasites at Different Concentrations.

Family	Plant	Active compound	Parasitic form	Concent- ration	Reference
Lamiaceae	Lavandula stoechas L	Flavonoids, phenolic acids, dipropenes, triplepenes, tannins, materials as bitter, resins, saponin	Giardia lamblia	400 mg/ml	Vazini <i>etal</i> ., 2017
Apiaceae	Ferula assa- foetida	α-Pinene, β-pinene, sabinene, eremophilene, β-caryophyllene and himachalen-7- ol	Giardia	20 mg/ml	Rezaee- Manesh and Shirbazou, 2012
Asteraceae	Tanacetum parthenium	Parthenolide	Giardia	100 mg/ml	Elmi <i>et al.</i> , 2014
Asteraceae	Allium paradoxum	Limonen, Spathulenol, alpha-Bisabolol, Z-Nerolidol, n-Tricosane, n-Docosane	Giardia	100 mg/ml	Elmi <i>et al.</i> , 2014
Apiaceae	Carum copticum	Thymol, Trinin, Pinen and Myrcens	Giardia	8 mg/ml	Shahabi et al., 2008
Amaryllidaceae	Allium ascalonicum	Thymol and carvacrol	Giardia	0.2 mg/ml	Azadbakht <i>et</i> al., 2003
Verbenaceae	Lippia berlandieri	Tannin and Vernalin	Giardia	0.85 mg/ml	Ponce <i>et al.</i> , 1994

REFERENCES

- [1] Yahia, E.M. ed., 2017. Fruit and vegetable phytochemicals: Chemistry and human health, 2 Volumes. John Wiley & Sons. 2nd Edition, pp.115-124.
- [2] Shitan, N., 2016. Secondary metabolites in plants: transport and self-tolerance mechanisms. Bioscience, biotechnology, and biochemistry, 80(7), pp.1283-1293.

- [3] Bansal, V., Kumar, P., Tuteja, S.K., Siddiqui, M.W., PRASAD, K. and SANGWAN, R., 2017. Diverse utilization of plant-originated secondary metabolites. Plant Secondary Metabolites, Volume Three: Their Roles in Stress Eco-physiology, p.215.
- [4] Symeonidou, I., Bonos, E., Moustakidis, K., Florou-Paneri, P., Christaki, E. and Papazahariadou, M., 2018. Botanicals: a natural approach to control ascaridiosis in poultry. Journal of the Hellenic Veterinary Medical Society, 69(1), pp.711-722.
- [5] Chanda, S. and Ramachandra, T.V., 2019. A review on some Therapeutic aspects of Phytochemicals present in Medicinal plants. International Journal of Pharmacy & Life Sciences, 10(1).
- [6] Capela, R., Moreira, R. and Lopes, F., 2019. An overview of drug resistance in protozoal diseases. International journal of molecular sciences, 20(22), p.5748.
- [7] Shanan, S., Abd, H., Bayoumi, M., Saeed, A. and Sandström, G., 2015. Prevalence of protozoa species in drinking and environmental water sources in Sudan. BioMed research international, 2015.
- [8] Imam, T.S., 2009. The complexities in the classification of protozoa: a challenge to parasitologists. Bayero Journal of Pure and Applied Sciences, 2(2), pp.159-164.
- [9] Antonovics, J., Wilson, A.J., Forbes, M.R., Hauffe, H.C., Kallio, E.R., Leggett, H.C., Longdon, B., Okamura, B., Sait, S.M. and Webster, J.P., 2017. The evolution of transmission mode. Philosophical Transactions of the Royal Society B: Biological Sciences, 372(1719), p.20160083.
- [10] Gardner, T.B. and Hill, D.R., 2001. Treatment of giardiasis. Clinical microbiology reviews, 14(1), pp.114-128.
- [11] Agrawal, A.K., Tripathi, D.M., Sahai, R., Gupta, N., Saxena, R.P., Puri, A., Singh, M., Misra, R.N., Dubey, C.B. and Saxena, K.C., 1997. Management of Giardiasis by a herbal drug Pippali Rasayana': a clinical study. Journal of ethnopharmacology, 56(3), pp.233-236.
- [12] Pintong, A.R., Ruangsittichai, J., Ampawong, S., Thima, K., Sriwichai, P., Komalamisra, N. and Popruk, S., 2020. Efficacy of Ageratum conyzoides extracts against Giardia duodenalis trophozoites: an experimental study. BMC complementary medicine and therapies, 20(1), pp.1-9.
- [13] Costa, T.M., Tavares, L.B.B. and de Oliveira, D., 2016. Fungi as a source of natural coumarins production. Applied Microbiology and Biotechnology, 100, pp.6571-6584.
- [14] Anvikar, A.R., Shah, N., Dhariwal, A.C., Sonal, G.S., Pradhan, M.M., Ghosh, S.K. and Valecha, N., 2016. Epidemiology of Plasmodium vivax malaria in India. The American journal of tropical medicine and hygiene, 95(6 Suppl), p.108.
- [15] Bahekar, S. and Kale, R., 2013. Herbal plants used for the treatment of malaria-a literature review. Journal of pharmacognosy and Phytochemistry, 1(6), pp.141-146.
- [16] Willcox, M., 2011. Improved traditional phytomedicines in current use for the clinical treatment of malaria. Planta medica, 77(06), pp.662-671.
- [17] Efferth, T., Herrmann, F., Tahrani, A. and Wink, M., 2011. Cytotoxic activity of secondary metabolites derived from Artemisia annua L. towards cancer cells in comparison to its designated active constituent artemisinin. Phytomedicine, 18(11), pp.959-969.
- [18] Thomas, K. and Ying, W., 2007. Artemisinin—an innovative cornerstone for anti-malaria therapy. Natural Compounds as Drugs: Volume II, pp.383-422.
- [19] Wink, M., 2008. Evolutionary advantage and molecular modes of action of multi-component mixtures used in phytomedicine. Current drug metabolism, 9(10), pp.996-1009.
- [20] Hellmann, J.K., Münter, S., Wink, M. and Frischknecht, F., 2010. Synergistic and additive effects of epigallocatechin gallate and digitonin on Plasmodium sporozoite survival and motility. PloS one, 5(1), p.e8682.
- [21] Alebie, G., Urga, B. and Worku, A., 2017. Systematic review on traditional medicinal plants used for the treatment of malaria in Ethiopia: trends and perspectives. Malaria Journal, 16, pp.1-13.
- [22] Alnassan, A.A., Thabet, A., Daugschies, A. and Bangoura, B., 2015. In vitro efficacy of allicin on chicken Eimeria tenella sporozoites. Parasitology research, 114, pp.3913-3915.
- [23] El-Khtam, A. O., Abd El Latif, A., & El-Hewaity, M. H. (2014). Efficacy of turmeric (Curcuma longa) and garlic (Allium sativum) on Eimeria species in broilers. International Journal of Basic and Applied Sciences, 3(3), pp. 349–356, 2014.
- [24] M. Pourali, H. Kermanshahi, A. Golian, G. R. Razmi, and M. Soukhtanloo, "Antioxidant and anticoccidial effects of garlic powder and sulfur amino acids on Eimeria-infected and uninfected broiler chickens," Iranian Journal of Veterinary Research, vol. 15, no. 3, pp. 227–232, 2014.
- [25] Shetshak, M.A., Suleiman, M.M., Jatau, I.D., Ameh, M.P. and Akefe, I.O., 2021. Anticoccidial efficacy of Garcinia kola (Heckel H.) against experimental Eimeria tenella infection in chicks. Journal of Parasitic Diseases, 45(4), pp.1034-1048.

- [26] Biswas, K., Chattopadhyay, I., Banerjee, R.K. and Bandyopadhyay, U., 2002. Biological activities and medicinal properties of neem (Azadirachta indica). Current science, pp.1336-1345.
- [27] Koul, A., Ghara, A.R. and Gangar, S.C., 2006. Chemomodulatory effects of Azadirachta indica on the hepatic status of skin tumor bearing mice. Phytotherapy Research: An International Journal Devoted to Pharmacological and Toxicological Evaluation of Natural Product Derivatives, 20(3), pp.169-177.
- [28] Tipu, M.A., Pasha, T.N. and Ali, Z., 2002. Comparative efficacy of salinomycin sodium and neem fruit (Azadirachta indica) as feed additive anticoccidials in broilers. International Journal of Poultry Science, 1(4), pp.91-93.
- [29] Rhayour, K., Bouchikhi, T., Tantaoui-Elaraki, A., Sendide, K. and Remmal, A., 2003. The mechanism of bactericidal action of oregano and clove essential oils and of their phenolic major components on Escherichia coli and Bacillus subtilis. Journal of essential oil research, 15(5), pp.356-362.
- [30] Chami, F., Chami, N., Bennis, S., Trouillas, J. and Remmal, A., 2004. Evaluation of carvacrol and eugenol as prophylaxis and treatment of vaginal candidiasis in an immunosuppressed rat model. Journal of antimicrobial chemotherapy, 54(5), pp.909-914.
- [31] Chami, F., Chami, N., Bennis, S., Bouchikhi, T. and Remmal, A., 2005. Oregano and clove essential oils induce surface alterat ion of Saccharomyces cerevisiae. Phytotherapy Research: An International Journal Devoted to Pharmacological and Toxicological Evaluation of Natural Product Derivatives, 19(5), pp.405-408
- [32] Bennis, S., Chami, F., Chami, N., Bouchikhi, T. and Remmal, A., 2004. Surface alteration of Saccharomyces cerevisiae induced by thymol and eugenol. Letters in Applied Microbiology, 38(6), pp.454-458.
- [33] Remmal, A., Achahbar, S., Bouddine, L., Chami, N. and Chami, F., 2011. In vitro destruction of Eimeria oocysts by essential oils. Veterinary parasitology, 182(2-4), pp.121-126.
- [34] Gehrig, S. and Efferth, T., 2009. Development of drug resistance in Trichomonas vaginalis and its overcoming with natural products. The Open Bioactive Compounds Journal, 2(1), pp.21-28.
- [35] Lourens, G.B. and Ferrell, D.K., 2019. Lymphatic filariasis. Nursing Clinics, 54(2), pp.181-192.
- [36] Singh, N.P.; Singh, V.K. Streblusasper—An ancient Indian drug for the cure of filariasis. Acta Bot. Indica 1987, 15, pp.108–109.
- [37] Chatterjee, R.K., Fatma, N., Murthy, P.K., Sinha, P., Kulshrestha, D.K. and Dhawan, B.N., 1992. Macrofilaricidal activity of the stembark of Streblus asper and its major active constituents. Drug Development Research, 26(1), pp.67-78.
- [38] Ibrahim, A.Y., Mahmoud, M.G. and Asker, M.M., 2014. Anti-inflammatory and antioxidant activities of polysaccharide from Adansonia digitata: An in vitro study. Int J Pharm Sci Rev Res, 25(2), pp.174-182.
- [39] Bazh, E.K. and El-Bahy, N.M., 2013. In vitro and in vivo screening of anthelmintic activity of ginger and curcumin on Ascaridia galli. Parasitology research, 112(11), pp.3679-3686.
- [40] Yazwinski, T.A., Tucker, C.A., Wray, E., Jones, L. and Clark, F.D., 2013. Observations of benzimidazole efficacies against Ascaridia dissimilis, Ascaridia galli, and Heterakis gallinarum in naturally infected poultry. Journal of Applied Poultry Research, 22(1), pp.75-79.
- [41] Umar, S., Abbas, S., Irfan Khan, M., Nisa, Q.U., Younus, M., Aqil, K., Qayyum, R., Yaqoob, M., Ali, A., Asif Yaseen, M. and Ali Shah, M., 2018. Safety of fenbendazole in common peafowl (Pavo cristatus). Pakistan Journal of Pharmaceutical Sciences, 31(1).
- [42] Zaman, M.A., Abbas, R.Z., Qamar, W., Qamar, M.F., Mehreen, U., Shahid, Z. and Kamran, M., 2020. Role of secondary metabolites of medicinal plants against Ascaridia galli. World's Poultry Science Journal, 76(3), pp.639-655.
- [43] Abdelqader, A., Qarallah, B., Al-Ramamneh, D. and Daş, G., 2012. Anthelmintic effects of citrus peels ethanolic extracts against Ascaridia galli. Veterinary Parasitology, 188(1-2), pp.78-84.
- [44] Mullner, A., Helfer, A., Kotlyar, D., Oswald, J. and Efferth, T., 2011. Chemistry and pharmacology of neglected helminthic diseases. Current medicinal chemistry, 18(5), pp.767-789.
- [45] Liu, S.Y., Sporer, F., Wink, M., Jourdane, J., Henning, R., Li, Y.L. and Ruppel, A., 1997. Anthraquinones in Rheum palmatum a nd Rumex dentatus (Polygonaceae), and phorbol esters in Jatropha curcas (Euphorbiaceae) with molluscicidal activity against the schistosome vector snails Oncomelania,
- [46] Biomphalaria, and Bulinus. Tropical Medicine & International Health, 2(2), pp.179-188.
- [47] Haddad, M., Sauvain, M. and Deharo, E., 2011. Curcuma as a parasiticidal agent: a review. Planta medica, 77(06), pp.672-678.
- [48] Ouedraogo, N., Atchade, C. and Kadiatou, T.T., 2020. Anti-Inflammatory activity of extracts from Parkia biglobosa (Jacq.) R. Br. Ex G. Don. (Fabaceae-Mimosoideae) trunk bark. J Pharmacol Toxicol, 16(1), pp.1-8.

- [49] Saleh, M.S., Jalil, J., Zainalabidin, S., Asmadi, A.Y., Mustafa, N.H. and Kamisah, Y., 2021. Genus Parkia: Phytochemical, medicinal uses, and pharmacological properties. International Journal of Molecular Sciences, 22(2), p.618.
- [50] Mukherjee, N., Joardar, N. and Babu, S.S., 2019. Antifilarial activity of azadirachtin fuelled through reactive oxygen species induced apoptosis: a thorough molecular study on Setaria cervi. Journal of helminthology, 93(5), pp.519-528.
- [51] Hamisi, M.M., Calista, I., Nteghenjwa, A.K., Shaaban, J.K., Richard, S., Frank, M., Patrick, K.T., Vitus, A.N., Victor, W., George, L. M. and John, W.O., 2013. Repellence effectiveness of essential oils from some Tanzanian Ocimum and Hyptis plant species against afro-tropical vectors of malaria and lymphatic filariasis. Journal of Medicinal Plants Research, 7(11), pp.653-660.
- [52] Vazini, H., 2017. The effect of lavender hydroalcoholic extract on cyst infection Giardia lamblia in mice. J. Neyshabour Univ. Med. Sci, 5(2), pp.22-31.
- [53] Rezaeemanesh, M., Shirbazoo, S. and Pouryaghoub, N., 2013. In-vitro giardicidal effects of aqueous and alcoholic extracts of Chenopodium botrys L. on Giardia lamblia cysts. Journal of Torbat Heydariyeh University of Medical Sciences, 1(1), pp.21-31.
- [54] Elmi, T., Gholami, S., Azadbakht, M., Rahimi-Esboei, B. and Geraili, Z., 2014. The effects of hydroalcoholic extract of leaves and onion of Allium paradoxum on Giardia lamblia in mice. Journal of Shahrekord University of Medical Sciences, 16(5), pp.13-22.
- [55] Shahabi, S., Ayazi Roozbehani, F., Kamalinejad, M. and Abadi, A., 2009. Anti-giardia activity of Carum copticum on Giardia lamblia cysts in vitro. Pejouhesh Dar Pezeshki, 32(4).
- [56] Azadbakht, M., Chabra, A., Saeedi Akbarabadi, A., Motazedian, M.H., Monadi, T. and Akbari, F., 2020. Anti-parasitic activity of some medicinal plants essential oils on Giardia lamblia and Entamoeba histolytica, in vitro. Res J Pharmacogn, 7(1), pp.41-47.
- [57] Ponce-Macotela, M., Navarro-Alegria, I., Martinez-Gordillo, M.N. and Alvarez-Chacon, R., 1994. In vitro effect against Giardia of 14 plant extracts. Revista de investigación clinica; organo del Hospital de Enfermedades de la Nutrición, 46(5), pp.343-347.