**A REVIEW ON BIOLOGY OF FALL ARMYWORM, *Spodoptera frugiperda* ON MAIZE**

**L. S. CHAUDHARY 1, C. J. PATEL 2, H. G. PATEL3 and D. S. RABARI 4**

1. **Department of Entomology, N M College of Agriculture, Navsari Agricultural University, Navsari- 396 450 Gujarat, India**

**Email address** **lalabhai261999@gmail.com** **and Mobile no. 8511421905**

1. **ASPEE Shakilam Biotechnology Institute, Navsari Agricultural University Surat-395 007 Gujarat, India**

**Email address** **cjpatel@nau.in** **and Mobile no. 9725006536**

1. **Department of Entomology, N M College of Agriculture, Navsari Agricultural University, Navsari- 396 450 Gujarat, India**

**Email address** **hiralpatel6078@gmail.com** **and Mobile no. 9723695936**

1. **Department of Entomology, N M College of Agriculture, Navsari Agricultural University, Navsari- 396 450 Gujarat, India**

**Email address** **dsrabari@nau.in** **and Mobile no. 8511680714**

**ABSTRACT**

The female laid eggs in masses (150-200 eggs/mass) on the under or upper surface of the maize leaf, lid of glass jar and also in whorls. The egg mass was covered with a layer of scales some time, not covered with scales. The incubation, larval, pre-pupal and pupal period were 2 to 3, 13.5 to 23, 2 to 3 and 5 to 8 days, respectively under laboratory condition. The first, second, third, fourth, fifth and sixth instar period were from 2 to 3, 2 to 3, 2 to 3, 2 to 3, 3 to 4 and 3 to 5 days respectively. The last instar larva was dark brown with reddish brown head marked with inverted ‘Y’ shape on the head with the elevated distinct dark coloured black spots (Pinacula) on the whole body which bears spines and arrangement of the dorsal pinacula the four black spots arranged in square on the 8th abdominal segment. The forewing of male adult was light brown, grey and straw. Markings on the male were more pronounced than the female with males having a grey colour and a light diagonal marking on the forewing. While female forewing was uniform greyish brown to a fine mottling of grey and brown. The life cycle of male and female were 28 to 41 days and 30 to 45 days, respectively.

**Keywords:** Fall armyworm, *Spodoptera frugiperda,* Maize

**INTRODUCTION**

The name “fall armyworm” originates from their nature of the damage, where infestations sometimes resemble as an army, as they move across large agricultural fields and earned their common name by eating all plant matter, they encounter in their wide dispersals, like a large army (Smith, 1797). Due to its migratory behaviour, the fall armyworm is known as a sporadic pest. A new invasive pest, fall armyworm (FAW), *Spodoptera frugiperda* (J. E. Smith) (Lepidoptera: Noctuidae) attained the status as a major pest resulting in extensive yield losses of maize all over the world. FAW is a devouring pest native to tropical and subtropical regions of America where it was first detected in 1797 and first discovered in the African continent in 2016 (Goergen *et al.,* 2016) and has reached China, spreading across Africa and Asia continents, west to east, in just three years. Entry of this destructive insect into a portion of Asia is questionable because a majority of people inhabit there and in locations nearby and already a huge pressure is created on food production systems. In India, the occurrence and prevalence of this invasive pest were noticed for the first time by Sharanabasappa *et al.* (2018) from Karnataka state which is the first reported infestation in Asia, in July 2018. Subsequently, it spread into the other ten states of India till the middle of March 2019. Later, its existence was confirmed in the states of Maharashtra, Gujarat and Chhattisgarh (Chormule *et al.*, 2019; Sisodiya *et al.,* 2018; Deole and Paul, 2018). It is a cosmopolitan pest of the maize crop (Wiseman *et al.*, 1966). The fall armyworm may travel over 500 km before they start oviposition (Prasanna *et al.*, 2018). The study of the biology of fall armyworm as occurring in Gujarat is very much important for identifying the life stages and finding out the weaker stage of the pest which provides the basis for management strategies. Farmers are growing a wide variety of commercial hybrids across the state, but the hybrids with good plant vigour and genetic resistance to crop pests were most preferred to combat the invasive alien insect pests. The use of insect-resistant cultivars is an important component of Integrated Pest Management (IPM) which provides an economic, stable and environmentally sound approach to minimize damage from borer pests (Rasool *et al.,* 2017). Keeping this in view present studies were carried out.

### **Egg Stage**

gravid females lay eggs in groups of lower or upper corn leaves, on the lids of glass jars and in rings. The eggs are dome-shaped, flat on the back and abdomen, first light green, then golden yellow, and finally black. Sometimes the eggs are covered with a layer of scales, while the large eggs found by Siddhapara *et al*. not covered with scales. (2021), Ramzan *et al*. (2021), Ahmed *et al.* (2021), Manjula *et al.* (2019), Sharanabasappa *et al*. (2018), Sisodia *et al*. (2018) and Rajisha *et al*. (2022). Rajisha *et al*. (2022) note that the eggs are placed as 150-200 eggs/group. The average fertility per female of 939 eggs was recorded by Vishwakarma *et al*. (2020). The egg incubation period is 2 to 3 days, according to Kranthi *et al.* (2022), Ashok *et al.* (2020), Vishwakarma *et al.* (2020), Ahmed *et al.* (2021), Sharanabasappa *et al.* (2018), Rajisha *et al.* (2022) and Kranthi *et al.* (2022). Kayan *et al.* (2020) noted that the incubation period of eggs is 3-4 days, with an average of 3.30 days. Egg diameters between 0.47 and 0.59 mm have been recorded by Manjula *et al.* (2019) and Ipsita and Sahu (2020).

### **Larva Stage**

#### **First instar**

Assefa (2018), Deole and Paul (2018), Sisodiya *et al.* (2018) It showed that the first star of FAW was black-headed green and Sharma *et al.* (2022) noticed the larvae are so small that they eat whole eggs. Kalyan *et al.* (2020) he noticed a large, flattened, round, black head and a white body covered with small hairs. The duration of first instar larvae ranged from 2.00-3.00 days with a mean of 2.6±0.51 days noted by Reddy *et al.* (2021) and more or less similar with Vishwakarma *et al.* (2022). The duration of first instar was 5 days recorded by Manjula *et al.* (2019). The body length was 1.68±0.00 mm and width was 0.14±0.26 mm of first instar larvae recorded Ahmad *et al.* (2021). The body Length and width of first instar larvae were 1.42±0.28 mm and 0.28±0.06 mm, respectively (Reddy *et al.*, 2021). Larva measured 0.71 mm in length and with 0.16 mm head capsule width recorded by Manjula *et al.* (2019). The mean head capsule width was 0.33 mm, whereas, body length and width were 1.67 mm and 0.33 mm respectively by reported by Vishwakarma *et al.* (2022).

#### **Second instar**

Second instar larvae have an amber head and a white to yellowish body with a brown tinge on the back. Kalyan *et al.* (2020) and Bawani *et al.* (2019) indicates that the body also has a free top line and bottom line at this level. Vishwakarma *et al.* (2020) they found that the number of hairs per segment varied between 2-4. The duration of the second instar larvae is 2.00-3.00 days with a mean of 2.7 ± 0.48 days reported by Reddy *et al* (2021). The length and width of the second instar larvae is noted by Vishwakarma *et al* was 3.81mm and width 1.14mm. The mean length and width of second instar larvae were 3.32±0.48 mm and width 0.69±0.04 mm, respectively noted by Reddy *et al.* (2021). Sharanabasappa *et al.* (2018) found the head capsule width of second instar larvae was 0.48±0.01 mm. The head capsule width was 3.5 to 4 mm whereas length was about 0.5 mm (Manjula *et al.*, 2019) and Sharanabasappa *et al.* (2018) recorded the larval head capsule width was 0.48±0.01 mm.

#### **Third instar**

The body color of third instar larvae varies significantly from light white to greenish brown. Kalyan *et al.* (2020) reported that dorsal and dorsal lower alba lines were clearly visible and black spots were evident. The third period is brown with two to three white ridge lines (Ahmad *et al.,* 2021; Sharma *et al.*, 2022). Features reported by Rajisha *et al.* (2022 it is an inverted Y-shaped yellow epithelial suture on the head. The third instar larval stage is 2.20 ± 0.41 days and Vinay *et al.* (2022) suggest that 41 days should be in the 2-3 day range. Vishwakarma *et al*. (2022) recorded as third instar larval stage 1.48 days by. The third instar larval time is 1.95 ± 0.20 days for sorghum hosts and 2.00 ± 0.05 days for maize hosts (Keerthi *et al.,* 2016). Body length and width are 6.68 ± 0.14 mm and 0.82 ± 0, respectively.03 mm were recorded by Siddhapara *et al*., 2021, Sharma *et al*. (2022) and Vishwakarma *et al*. (2022). The average length of the third semester is 7.12 mm (6.4-7.8 mm), head capsule width 0.8 mm (0.80 ± 0.8 mm). 004) Sharma *et al.* (2022). Head capsule width is 0.50 ± 0.01 mm (Ahmad *et al*., 2017). Ib., 2021). The third instar larval duration was 1.48 days recorded by Vishwakarma *et al*. (2022). the 3rd instar larval period of 1.95±0.20 days on the sorghum host and 2.00±0.05 days on the maize hosts (Keerthi *et al*., 2021). The body length and width were 6.68±0.14 mm and 0.82±0.03 mm, respectively recorded by Siddhapara *et al*. (2021), Sharma *et al*. (2022) and Vishwakarma *et al*. (2022). The average length of 3rd instar was 7.12 mm (6.4-7.8 mm) and width of head capsule was 0.8 mm (0.80±0.004) recorded by Sharma *et al*. (2022). The head capsule width was 0.50±0.01 mm (Ahmad *et al*., 2021).

Fourth instar

The body color of the fourth instar larvae varies from olive brown to dark brown. Kalyan *et al*. (2020) found that the dorsal and dorsal inferior linea alba also appeared. Reddy *et al*. (2020) found a clearly seen inverted "Y" shaped mark on the head and four black dots arranged in a square on 8th abdominal segment. All the spots arranged as in case of third instar with increase in size and with more darkness reported by Manjula *et al.* (2019). The duration of fourth instar larvae ranged from 2.00-3.00 days with a mean of 2.4±0.51 days noted by Reddy *et al. (*2020) and Reddy *et al.* (2021). The body length and width were measured 13.94±1.09 mm and 0.84±0.02 mm, respectively recorded by Siddhapara *et al. (*2021). The average length was 11.6 mm (9.4-13.5 mm) and width of head capsule was 1.37±0.006 mm measured by Sharma *et al., (*2022).

Fifth instar:

Fifth instar larvae are pale yellow to cream in colour. Head colour is brick red with dark brown spots. The colour of the chest and abdomen is similar. Only the area around the first chest hair is a black ring. There are also dark circles and lighter areas.The shallow dorsal longitudinal line will be shallow. Manjula *et al*.(2019) and Igyuve *et al*. (2018). see the brown color in the outer area. Spiny black nodules were found on the dorsal side of the body (Reddy *et al*., 2020). The duration of fifth instar larva varied from 3 to 4 days with an average of 3.05±0.05 days recorded by Tiwari and Deole (2021); Rajisha *et al.* (2022) and Reddy *et al.* (2020). The mean body length and width were 18.81 mm and 3.45 mm, respectively (Vishwakarma *et al.*, 2022). The average length of 5th instar was 18.51 mm (15.8-22.5 mm) and width of head capsule was 2.11 mm reported by Sharma *et al. (*2022). Vishwakarma *et al.* (2022) recorded the mean body length and width were 18.81 mm and 3.45 mm, respectively.

Sixth instar:

The sixth instar larva as dark brown with reddish brown head marked with inverted ‘Y’ shape on the head with the elevated distinct dark coloured black spots (Pinacula) on the whole body which bears spines (long primary setae). The arrangement of the dorsal pinacula the four black spots arranged in square on the 8th abdominal segment. Large spots especially on 9th segment had a typical arrangement in a trapezoidal pattern and also seen from 1 to 7th abdominal segments (Babu *et al.,* 2019; Sharanabasappa *et al.*, 2018; Manjula *et al.*, 2019; Reddy *et al.*, 2021 and Vinay *et al.,* 2022). The duration of 3.67±0.83 days of sixth instar larval period which varied from 3-5 days reported by Reddy *et al. (*2020) and Vishwakarma *et al.* (2022). The mean length and width of sixth instar larvae were 33.6±1.67 mm and 5.90±0.26 mm, respectively noticed by Reddy *et al.*, (2021) and Vishwakarma *et al.* (2022). Ahmad *et al.* (2021) reported that length and width of larva were 26.98±2.93 and 3.90±0.00 mm, respectively. The average length of 6th instar larva was 34.39 mm (28.3-38.8 mm) and width of head capsule was 2.7 mm noted by Sharma *et al. (*2022).

#### **Total larval period**

The total larval period varied from 13.5 to 23 days with an average of 18.25 days recorded by Vishwakarma *et al. (*2020). Sharanabasappa *et al.* (2018); Bhavani *et al.* (2019); Manjula *et al.* (2019) and Vishwakarma *et al.* (2020) noted the total larval duration was 14-19, 13-14, 14-30 and 13.5 to 23 days, respectively.

Pre pupal stage:

Sharanabasappa *et al*. (2018) Bawani *et al*. (2019) and Vishwakarma *et al*. (2022) They found that before the pupal stage, the mature larvae stop eating and turn green and bright brown. The incubation period is 2 to 3 days, with an average of 2 to 3 days. with an average of 2.10±0.06 days recorded by Tiwari and Deole (2021 and Kranthi *et al.,* 2022). The length of the pre-pupae ranging from 19 to 22 mm with an average of 20.0±0.11 mm while the breadth ranged from 2.7 to 3.2 mm with an average of 3.08±0.02 mm observed by Tiwari (2020).

### **Pupa Stage**

Newly formed pupae are green in color. After 12-14 hours, the color of the fry changed to a deep red color as described by Deole and Paul (2018). Ahmed *et al*. (2021) It has been observed that pupae occur on corn stalks or cut leaves or on the rim of petri dishes. Sisodia *et al*. (2018) and Babb *et al*. (2019) noted a reddish-brown coloration in the custard in pupae. Ahmed *et al*. (2021) mentioned an important morphological feature such as the purpose of pupae being white-green. The education period of primary school students is between 5 and 8 days, with an average of average of 6.30±0.14 days (Tiwari and Deole, 2021). ). Sharanabasappa *et al.* (2018) recorded the pupal duration period was 9 to 12 days. The distance between genital opening and anal slot recorded more in case of female (0.89±0.01 mm) than the male (0.43±0.01 mm) recoded by Siddhapara *et al.* (2021). Length and width of male and female pupa were 14.12±0.38, 14.00±0.32 and 4.12±0.12, 4.00±0.00, respectively (Ahmad *et al.*, 2021 and Vishwakarma *et al.*, 2022)). The average weight of pupa was 18.9 mg (15-24 mg) weighted by Sharma *et al.* (2022). Gamil (2020) recorded 9.56 days pupal duration and the pupal weight was 0.3033 gm, while the normal pupae, malformed pupae, pupal mortality and emergence per cent was 95.44, 4.56, 4.0, and 96.0 per cent, respectively.

**Adult Stage**

#### **Male**

Adult male fore wings are light brown, gray and straw colored. The markings are more pronounced in males than females, and males are gray with a slight diagonal on the fore wings. Deole and Paul (2018) concluded that the hind wings are white. Sisodia *et al*. (2018) shows gray-brown male adults. The forewings are shades of gray and brown with oval or oblique orbital spots and triangular white spots near the apical edge of the forewings. Damasi *et al*. (2020) found that the adult male is gray-brown in color, with shades of gray and brown on the forewings, an oval orbital point and a triangular white spot near the apical edge of the forewings. Sharanabasappa *et al*. (2018) reported that adult males live 8.20 days, with a range of 7-9 days and an average wingspan of 3. 25 cm long with range of 3.00 to 3.50 cm in male. The average body length of male moths is 15.20 ± 1.30mm (Reddy *et al*., 2021).

#### **Female Adult**

The forewings of females were uniform greyish brown to a fine mottling of grey and brown. The hind wing was silver-white with a narrow dark border in both male and female (Sharanabasappa *et al.*, 2018; Reddy *et al.*, 2021 and Sharma *et al.,* 2022). The female body length was 10.13±0.56 mm and wing expanse was 31.00 ±1.43 mm measured by Ahmad *et al.* (2021). The female adult survived for 9-12 days with an average of 10.80 days (Sharanabasappa *et al.*, 2018). Manjula *et al.* (2019) revealed 1.2 cm forewing length, 0.9 cm hind wing length and 3.0 to 3.1 cm wing span. Ahmad *et al.* (2021) reported that the female body length was 10.13±0.56 mm and wing expanse was 31.00 ±1.43 mm. The average weight of adult female 3.3 mg and wingspan of female was measured as 3.2 cm reported by Sharma *et al.,* 2022).

### **Pre-oviposition, Oviposition and Post-oviposition Period**

The pre-oviposition period of female moths of *S. frugiperda* varied from 3 to 4 days with an average of 3.05±0.05 days, however oviposition period was 1 to 2 days with an average of 1.85±0.08 days reported by Tiwari and Deole (2021). The post-ovipositional period ranged from 4 to 6 days with an average of 4.40±0.51 days recorded by Reddy *et al.* (2021). Gamil (2020) stated that the mean time required for maturation of the ovaries and starting to egg-laying (pre-oviposition period) was 3.50 days. Moreover, oviposition and post-oviposition period were 5.11 and 2.61 days, respectively. Vishwakarma *et al.* (2020) observed the mean pre-oviposition, oviposition and post-oviposition period as 3.5, 3.5 and 3.5 days and it ranged from 3-4, 2-5 and 3-4 days, respectively. Keerthi *et al.* (2021) revealed that the pre-oviposition period on sorghum and maize was 3.87±0.52 and 3.71±0.45 days, respectively whereas, oviposition period on sorghum and maize was 3.10±0.62 and 3.05±0.52 days. Siddhapara *et al.* (2021) noted the pre-oviposition, oviposition and post oviposition period was of 3.50±0.51, 3.20±0.89 and 3.90±0.71 days, respectively.

### **Life Span**

The total life cycle of *S. frugiperda* occupied on an average of 33.1±0.69 days ranging from 28 to 41 days in case of male, while 36.0±0.75 days ranging from 30 to 45 days in case of female observed by Tiwari and Deole (2021). Vinay *et al.* (2022) reported 38.10±6.51 days total duration of fall armyworm life cycle. Ahmad *et al.* (2021) noticed 35.32±4.02 and 42.00±5.76 days for average total life cycle of male and female, respectively. Sharanabasappa *et al.* (2018) reported total life cycle of male and female ranging from 32-43 and 34-46 days, respectively. Kalyan *et al.* (2020) revealed 36.15 and 40.11 days to complete total life cycle of male and female, respectively.

### **Sex Ratio**

The sex ratio of fall armyworm at constant conditions in the laboratory was approximately 1:1.23 (45.24 male:55.76 female) recorded by Gamil (2020). Siddhapara *et al.* (2021) calculated the sex ratio which was 1.13:1 (female: male). Sharma *et al.* (2022) worked out the adult male to female sex ratio as 1:1.30.

### **Fecundity**

The fecundity of 1004.65±110.00 eggs and range were 820-1150 eggs/female recorded by Ahmad *et al.* (2021). Reddy *et al.* (2021) reported that the egg laying capacity of female varied from 855-1172 eggs with an average of 1015±115.48 eggs. Sharanabasappa *et al.* (2018) reported that the average fecundity/female was 1064.80±109.53 eggs which ranged from 835-1169 eggs. Siddhapara *et al.* (2021) noticed 1145.43±182.15 eggs/female (766 to 1389 eggs). Tiwari and Deole (2021) noticed that egg laying capacity of female varied from 536 to 579 eggs with an average of 557.2±2.81 eggs. Sharma *et al.* (2022) revealed 979.43±24.086 eggs average fecundity and the range was 713 to 1166 eggs.

### **Hatchability**

The extent of egg hatching in the rage of 72.00-95.54 per cent reported by Siddhapara *et al.* (2021). Vinay *et al.* (2022) found the hatching of eggs was about 96.26 per cent. Sharanabasappa *et al.* (2018) reported 96.60±1.43 percent egg hatchability which ranged from 95-98 per cent. Gamil (2020) recorded the hatchability in fall armyworm and it was observed 97.33 per cent.

**CONCLUSION**

 The gravid female laid eggs in 150-200 eggs/mass on the under or upper surface of the maize leaf, lid of glass jar and also in whorls and egg mass covered with a layer of scales some time, not covered with scales. The incubation, larval, pre-pupal and pupal period were 2 to 3, 13.5 to 23, 2 to 3 and 5 to 8 days, respectively under laboratory condition. The newly formed pupae were green in colour. After 12-14 hours pupae were converted dark reddish brown in colour, The forewing of male adult was light brown, grey and straw. While female forewing was uniform greyish brown to a fine mottling of grey and brown. The hind wing of adults was whitish colour.

**REFERENCE**

A. Chormule, N. Shejawal, Sharanabassappa, C. M. Kalleshwaraswamy, R. Asokan and H. M. Mahadeva Swamy “First report of the fall armyworm, *Spodoptera frugiperda* (J. E. Smith) (Noctuidae: Lepidoptera) on sugarcane and other crops from Maharashtra, India” *Journal of Entomology and Zoology Studies*, 7(1): 114-117, 2019.

B. Bhavani, V. Chandra Sekhar, V. P. Kishore, L. M. Bharatha, P. Jamuna and B. Swapna “Morphological and molecular identification of an invasive insect pest, fall armyworm, *Spodoptera frugiperda* occurring on sugarcane in Andhra Pradesh, India” *Journal of Entomology and Zoology Studies*, 7(4): 12-18, 2019.

B. M. Prasanna, J. E. Huesing, R. Eddy and V. M. Peschke “Fall Armyworm in Africa: *A Guide for Integrated Pest Management*, First Edition” Mexico, CDMX: CIMMYT, pp. 13-14, 2018.

B. R. Wiseman, R. H. Painter and C. E. Wasson “Detecting corn seedling differences in the greenhouse by visual classification of damage by the fall armyworm” *Journal of Economic Entomology*, 59: 1211-1214, 1966.

D. B. Sisodiya, B. L. Raghunandan, N. A. Bhatt, H. S. Verma, C. P. Shewale, B. G. Timbadiya, and P. K. Borad “The fall armyworm, *Spodoptera frugiperda* (J. E. Smith) (Lepidoptera: Noctuidae); first report of new invasive pest in maize fields of Gujarat, India” *Journal of Entomology and Zoology Studies,* 6(5): 2089-2091, 2018.

D. Kalyan, M. K. Mahla, R. Babu, R. K. Kalyan and P. Swathi “Biological Parameters of *Spodoptera frugiperda* (J. E. Smith) under laboratory conditions” *International Journal of Current Microbiology and Applied Sciences,* 9(5): 2972-2979, 2020.

D. M. Damasia, J. J. Pastagia and H. R. Kachela “First Report of the occurrence of fall armyworm, *Spodoptera frugiperda* (J. E. Smith) on finger millet (*Eleusine coracana* Gaertn) in Gujarat, India” *International Journal of Current Microbiology and Applied Sciences,* Special Issue-11: 3608-3612, 2020.

F. Assefa “Status of fall armyworm (*Spodoptera frugiperda*), biology and control measures on maize crop in Ethiopia: a review”International Journal of Entomology Research, 06(02): 75-85, 2018.

G. Goergen, P. L. Kumar, S. B. Sankung, A. Togola and M. Tamo “First report of outbreaks of the fall armyworm *Spodoptera frugiperda* (J. E. Smith) (Lepidoptera, Noctuidae), a new alien invasive pest in west and central Africa” *Plos One,* 11(10), 2016.

I. Rasool, A. R.Wani, M. Nisar, Z. A. Dar, R. K. Nehru and B. Hussain “Antixenosis and antibiosis as a resistance mechanism to *Chilo partellus* (Swinhoe) (Lepidoptera: Pyralidae) in some maize genotypes*” Journal of Entomology and Zoology*, 5(2): 22-27, 2017.

J. E. Smith “First record of the fall armyworm, *Spodoptera frugiperda* (Lepidoptera: Noctuidae)” *African Entomology,* 26(1): 244-246, 1797.

K. Ashok, J. S. Kennedy, V. Geethalakshmi, P. Jeyakumar, N. Sathiah and V. Balasubramani “Life table study of fall armyworm *Spodoptera frugiperda* (J. E. Smith) on maize” *Indian Journal of Entomology* 82(3): 574-579, 2020.

K. J. M. Reddy, K. Kumari, T. Saha, and S. N. Singh “First record, seasonal incidence and life cycle of fall armyworm, *Spodoptera frugiperda* (J. E. Smith) in maize at Sabour, Bhagalpur, Bihar” *Journal of Entomology and Zoology Studies*, 8(5): 1631-1635, 2020.

K. Manjula, Y. Peeru Saheb, M. J. Sudheer and A. R. Rao “Studies on biology, feeding habits and natural enemies of fall armyworm, *Spodoptera frugiperda*, a new invasive pest in India” *Journal of Entomology and Zoology Studies*, 7(6): 1245-1250, 2019.

M. R. Siddhapara, K. M. Patel and A. G. Patel. “Biology and morphometrics of fall armyworm *Spodoptera frugiperda* (J. E. Smith) on maize” *Indian Journal of Entomology,* 83: pp 1-3, 2021.

M. Ramzan, H. Ilahi, M. Adnan, A. Ullah and A. Ullah “Observation on fall armyworm, *Spodoptera frugiperda* (Lepidoptera: Noctuidae) on maize under laboratory conditions” *Egyptian Academic Journal of Biological Sciences,* 14(1): 99-104, 2021.

N. A. Reddy, Y. S. Saindane, C. S. Chaudhari, and S. A. Landage “Biology of fall armyworm *Spodoptera frugiperda* (J. E. Smith) on maize under laboratory conditions” *The Pharma Innovation Journal*, 10(9): 1997-2001, 2021.

N. Vinay, S. V. S. Raju, S. R. Babu and K. R. Sharma “First report on occurrence of fall armyworm, *Spodoptera frugiperda* (J. E. Smith) (Lepidoptera: Noctuidae) from North Eastern Plain zone of Uttar Pradesh, India” *Journal of Experimental Zoology, India*, 25: 2059-2064, 2022.

P. Kranthi, R. Sunitha Devi and P. Rajanikanth “Biology of fall armyworm, *Spodoptera frugiperda* (J. E. Smith) (Lepidoptera: Noctuidae) on sugarcane” *Environment and Ecology,* 39(2): 442-446, 2022.

P. Kranthi, R. Sunitha Devi and P. Rajanikanth “Biology of fall armyworm, *Spodoptera frugiperda* (J. E. Smith) (Lepidoptera: Noctuidae) on sugarcane” *Environment and Ecology,* 39(2): 442-446, 2022.

P. S. Rajisha, N. Muthukrishnan, J. Nelson, R. Jerlin, P. Marimuthu and R. Karthikeyan “Biology and nutritional indices of the fall armyworm *Spodoptera frugiperda* (J. E. Smith) on maize” *Indian Journal of Entomology,* 84(1): 92-96, 2022.

R. Babu, R. K. Kalyan, S. Joshi, C. M. Balai, M. K. Mahla and P. Rokadia “Report of an exotic invasive pest the fall armyworm, *Spodoptera frugiperda* (J. E. Smith) on maize in Southern Rajasthan” *Journal of Entomology and Zoology Studies*, 7(3): 1296-1300, 2019.

R. Vishwakarma, P. Kumari, S. Patidar, S. B. Dasand, A. Nema, “First report of fall armyworm, *Spodoptera frugiperda* (J. E. Smith) (Lepidoptera: Noctuidae) on maize (*Zea mays*) from Madhya Pradesh, India*” Journal of Entomology and Zoology Studies*, 8(6): 819-823, 2020.

R. Vishwakarma, S. B. Das, S. Patidar, S. Mohanta, and V. K. Paradkar “Developmental and morphometric parameters of *Spodoptera frugiperda* on maize genotypes” *The Pharma Innovation Journal*, 11(9): 617-622, 2022.

S. Deole and N. Paul “First report of fall armyworm, *Spodoptera frugiperda* (J. E. Smith), their nature of damage and biology on maize crop at Raipur, Chhattisgarh” *Journal of Entomology and Zoology Studies,* 6(6): 219-221, 2018.

S. Ipsita and B. K. Sahu “Fall armyworm: Current status and management in India” *Journal of Entomology and Zoology Studies*, 8(3): 330-334, 2020.

S. Sharma, S. Tiwari, R. B. Thapa, S. Neupane, G. V. P. Reddy, S. Pokhrel and R. Muniappan, “Life cycle and morphometrics of fall armyworm (*Spodoptera frugiperda*) (Lepidoptera: Noctuidae) on maize crop” *SAARC Journal of Agriculture,* 20(1): 77-86, 2022.

S. Tiwari “Biology and behavioural studies of fall armyworm, *Spodoptera frugiperda* (J. E. Smith) (Lepidoptera: Noctuidae) along with its biorational management on *Kharif* maize crop” *M. Sc. Thesis*, Indira Gandhi Krishi Vishwavidyalaya, Raipur, pp. 62, 2020.

S. Tiwari and S. Deole “Studies on life cycle of fall armyworm, *Spodoptera frugiperda* (J. E. Smith) on maize at Raipur, Chhattisgarh” *The Pharma Innovation Journal,* 10(2): 643-646, 2021.

Sharanabasappa, C. M. Kalleshwaraswamy, M. S. Maruthi and H. B. Pavithra “Biology of invasive fall armyworm, *Spodoptera frugiperda* (J. E. Smith) (Lepidoptera: Noctuidae) on maize” *Indian Journal of Entomology,* 80(3): 540-543, 2018.

T. Ahmad, H. A. Ali, A. Ghaffar, K. Jehan, M. U. Mustafa, R. Ali, A. Faraz, and M. Ramzan “Biomorphic characters of fall armyworm, *Spodoptera frugiperda* (Lepidoptera: Noctuidae) on maize in Pakistan” *Egyptian Academic Journal of Biological Sciences*, 14(2): 13-18, 2021.

T. M. Igyuve, G. O. S. Ojo, M. S. Ugbaal and A. E. Ochigbo “Fall armyworm (*Spodoptera frugiperda*); it’s biology, impact and control on maize production in Nigeria” *Nigerian Journal of Crop Science,* 5(1): pp. 70-79, 2018.

W. E. Gamil “Fall armyworm *Spodoptera frugiperda* (J. E. Smith) (Lepidoptera: Noctuidae) biological aspects as a new alien invasive pest in Egypt” *Egyptian Academic Journal of Biological Sciences,* 13(3): 189-196, 2020.